Module Leader

Prof G Malliaras [1]

Lecturer

Prof G Malliaras & Prof E Hall [2]

Timing and Structure

Lent term. Lectures and coursework. Assessment: 100% coursework.

Aims

The aims of the course are to:

- Link engineering principles to understanding of biosystems in sensors and bioelectronics.

Objectives

As specific objectives, by the end of the course students should be able to:

- Extend principles of engineering to the development of biosensors and bioelectronic devices.
- Understand the principles of signal transduction between biology and electronics. • appreciate the basic configuration and distinction among biosensors and bioelectronic systems.
- Appreciate the basic configuration and distinction among biosensors and bioelectronic systems.
- Demonstrate appreciation for the technical limits of performance.
- Make design and selection decisions in response to measurement and actuation problems amenable to the use of biosensors and bioelectronic devices.
- Be able to evaluate novel trends in the field.

Content

This course covers the principles, technologies, methods and applications of biosensors and bioelectronics. The objective of this course is to link engineering principles to understanding these biosystems. It will provide the student with detail of methods and procedures used in the design, fabrication and application of biosensors and bioelectronic devices. The fundamentals of measurement science are applied to electrochemical and optical signal transduction. The fundamentals of electrophysiology are applied to implantable and cutaneous bioelectronic devices. Upon successful completion of this course, students are expected to be able to explain biosensing and transduction techniques, design and construct biosensor instrumentation, and explain the techniques of recording and stimulation of electrically active cells and tissues.

Introduction to Biosensors

- Overview of Biosensors
- Fundamental elements of biosensor devices
- Engineering sensor proteins
Electrochemical Biosensors

- Electrochemical principles
- Amperometric biosensors and charge transfer pathways in enzymes
- Glucose biosensors
- Engineering electrochemical biosensors

Optical Biosensors

- Optics for biosensors
- Attenuated total reflection systems

Diagnostics for the real world

- Communication and tracking in health monitoring
- Detection in resource limited settings

Introduction to Bioelectronics

- Overview of technology (implantable, cutaneous, ex vivo)
- Anatomy, function of nervous system, other electrically active tissues
- Principles of electrophysiology
- Recording and stimulation (intracellular, extracellular, epidural, EEG)
- Transducers (pipette electrodes, Ag/AgCl, metal electrodes, Michigan and Utah probes, transistors)

Implantable devices

- Cardiac pacemaker
- Cochlear implant, retinal implant
- DBS (Parkinson’s, dystonia, epilepsy), spinal cord stimulators
- Brain-Computer Interfaces
- PNS stimulators, electroceuticals
- Implantable drug delivery systems
- Foreign body reaction

Wearable devices

- Cutaneous electrophysiology (brain, heart, muscle)
- Electronic skins (pressure, temperature)
- Sweat biosensing (glucose, lactate, …)
- Transdermal drug delivery

Ex vivo devices

- Electrochemical biosensors
- Impedance biosensors
- MEAs and patch clamp
- Organ-on-a-chip
- In vitro systems

Regulatory and ethical issues

Coursework

The coursework will be assessed on two marked assignments. The first assignment will involve a demonstration of glucose sensor technology. The second assignment will involve an illustration of the principles of electrophysiology applied to bioelectronic devices.
### Coursework

<table>
<thead>
<tr>
<th>Coursework activity #1 : Glucose biosensors</th>
<th>Format</th>
<th>Due date &amp; marks</th>
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<tbody>
<tr>
<td>Learning objectives:</td>
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<tr>
<td>• To introduce students to electrochemical sensors employed for the measurement of glucose.</td>
<td>Individual Report</td>
<td>17th Feb 21 [30/60]</td>
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<td>• To quantitatively analyse measurements conducted using test strip glucose biosensors on a range of samples.</td>
<td>anonymously marked</td>
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<td>• To explore the principles of accuracy and precision and their application to clinical measurements and their impact on clinical decision making.</td>
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<td>• To extend the principles to the design of a biosensor for the measurement of lactate.</td>
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<tr>
<th>Coursework activity #2 : Cutaneous electrophysiology</th>
<th>Format</th>
<th>Due date &amp; marks</th>
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<tr>
<td>Learning objectives:</td>
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<tr>
<td>• To introduce students to sensors employed for the measurement of electrophysiology.</td>
<td>Individual Report</td>
<td>17th March 21 [30/60]</td>
</tr>
<tr>
<td>• To explore different recording configurations.</td>
<td>anonymously marked</td>
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<tr>
<td>• To quantitatively analyse measurements conducted using cutaneous electrodes.</td>
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<tr>
<td>• To extend the principles to the design of a sensor for the measurement of biopotentials.</td>
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### Booklists

Please refer to the Booklist for Part IIB Courses for references to this module, this can be found on the associated Moodle course.

### Examination Guidelines

Please refer to Form & conduct of the examinations [3].

### UK-SPEC

The [UK Standard for Professional Engineering Competence (UK-SPEC)](https://www.engc.org.uk/standards/uk-spec) [4] describes the requirements that have to be met in order to become a Chartered Engineer, and gives examples of ways of doing this.

UK-SPEC is published by the Engineering Council on behalf of the UK engineering profession. The standard has been developed, and is regularly updated, by panels representing professional engineering institutions, employers and engineering educators. Of particular relevance here is the ‘Accreditation of Higher Education Programmes (AHEP)’ document [5] which sets out the standard for degree accreditation.


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Links
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