

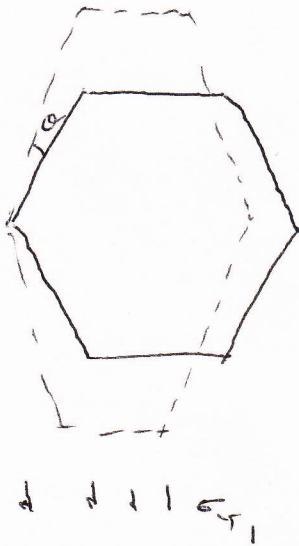
(a)

Honeycomb : 
$$\frac{3 t_1 l_1}{2 \left[ \frac{1}{2} \sqrt{3} l_1 \frac{l_1}{2} \right] + \sqrt{3} l_1^2} = \frac{2 t_1}{\sqrt{3} l_1} = \bar{P}_1$$

Triangular : 
$$\frac{3 t_2 l_2}{\frac{\sqrt{3}}{2} l_2^2} = 2 \sqrt{3} \frac{t_2}{l_2} = \bar{P}_2$$

$\uparrow \sigma_{T1}, \epsilon$

(b)



$$\sigma_{T1} \epsilon \frac{3}{2} \sqrt{3} l_1^2 = \frac{M_p (4\theta \times 2)}{2}$$

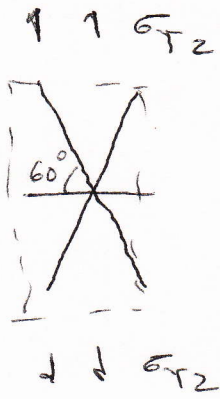
$$M_p = \frac{\sigma_{TS} t_1^2}{4}$$

$$\epsilon = \frac{l_1 \theta \sin 30^\circ}{l_1 \sin 60^\circ} = \frac{\theta}{\sqrt{3}}$$

$$\sigma_{T1} \frac{\theta}{\sqrt{3}} \frac{3 \sqrt{3} l_1^2}{2} = \frac{\sigma_{TS} t_1^2}{4} 4 \theta$$

$$\sigma_{T1} = \sigma_{TS} \frac{2}{3} \left( \frac{t_1}{l_1} \right)^2 = \frac{\bar{P}_1^2}{2} \sigma_{TS}$$

(c)



$$\sigma_F t_2 \frac{\sqrt{3}}{2} \times 2 = \sigma_{T2} l_2$$

$$\sigma_{T2} = \sqrt{3} \left( \frac{t_2}{l_2} \right) \sigma_f$$

$$= \frac{\bar{P}_2}{2} \sigma_f$$

(d)

$$\sigma_{T5} = \sigma_f \frac{\bar{P}_2}{2}$$

$$\sigma_{T1} = \frac{\bar{P}_1^2 \bar{P}_2}{4} \sigma_f$$

2  
(a)

$$-v \frac{dn}{dx} = (1-n)f(x) - ng(x)$$

$$v=0$$

$$(1-n)f = ng$$

$$n = \frac{f}{f+g}$$

(b) For shooting ~~n=0~~  $n=0$   $x > h$  as no cross-bridges are dragged there & ~~f~~  $f=0$   
for  $x > h$

$$\underline{0 < x < h}$$

$$-v \frac{dn}{dx} = (1-n)f_0 - ng_0$$

$$n = \underbrace{A \exp\left[\frac{(f_0+g_0)x}{v}\right]}_{\text{homogeneous}} + \underbrace{\frac{f_0}{f_0+g_0}}_{\text{particular}}$$

$$n(h) = 0$$

$$\Rightarrow A = \frac{-f_0}{(f_0+g_0) \exp\left[\frac{(f_0+g_0)h}{v}\right]}$$

$$n(x) = \frac{f_0}{f_0+g_0} \left[ 1 - \exp\left[\frac{(f_0+g_0)}{v} (x-h)\right] \right]$$

$$\text{ie } n(0) = \frac{f_0}{f_0 + g_0} \left[ 1 - \exp \left\{ -\frac{(f_0 + g_0)h}{v} \right\} \right]$$

$$\underline{x < 0}$$

$$-v \frac{dn}{dz} = -ng_1$$

$$n = C \exp \left( \frac{g_1 x}{v} \right)$$

$$n(0) = C = \frac{f_0}{f_0 + g_0} \left[ 1 - \exp \left\{ -\frac{(f_0 + g_0)h}{v} \right\} \right]$$

$$n(x) = \frac{f_0}{f_0 + g_0} \left[ 1 - \exp \left\{ -\frac{(f_0 + g_0)h}{v} \right\} \right] \exp \left( \frac{g_1 x}{v} \right)$$

(c)

It would ~~not~~ involve writing a PDE for  $n(x,t)$  of the form

$$\frac{\partial n}{\partial t} = -v \frac{\partial n}{\partial x} = (1-n)f - ng$$

and solving subject to  $\frac{\partial T_1}{\partial t} = 0$ .

3(a)

(i) Persistence length is a material property dependent on bending stiffness  $D$  & thermal activation  $kT$

$$\lambda_p = \frac{D}{kT}$$

Over a length  $\lambda_p$  ~~angle~~ angular correlation is lost & the filament can be thought to be rubber-like. ~~Thus~~ Thus if filament length  $l \gg \lambda_p$  then modulus is entropic & if  $l \ll \lambda_p$  then fibre behaves like a stiff beam.

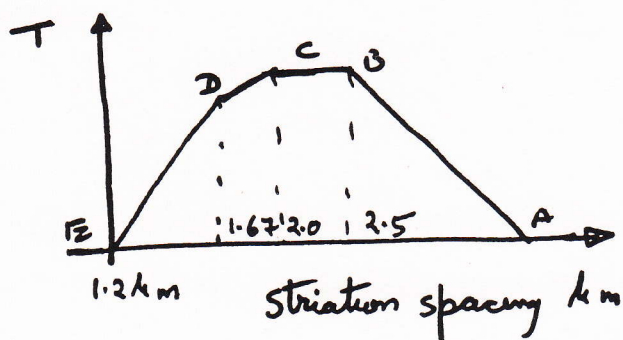
(ii) If nodal ~~con~~ connectivity is high (eg. triangulated in 2D) then, modulus of structure is governed by stretching stiffness ~~strength~~ of filaments but if connectivity is low (honeycombs in 2D) modulus is governed by bending stiffness of filaments

3 (b)

With a high  $Ca^{2+}$  concentration in the sarcoplasm, the myosin head is tightly bound to an adjacent actin filament. Before long ATP binds to the myosin & a conformational change reduces the affinity of myosin to actin & the two separate, simultaneously causing the myosin to shift a distance of 5nm towards the +ve end of the actin filament, where it rebinds at a new location. Hydrolysis then causes the release of  $P_i$  from the ATP to produce ADP & the associated conformational change triggers the power stroke that draws the actin filament in the direction of the -ve end. During the power stroke the ADP is released returning the myosin to its original state, ready for the next ATP to come along & bond.

(25%)

(c)



The tension changes with change in overlap between the thick & thin filaments. Between B & C the overlap is a constant & so is the tension.

4(a)

The cytoskeleton is the internal soft skeleton of the cells. The microtubules are stiff & hollow tubes made from tubulin. They emanate from the centrosome & small molecular motors carry proteins along them. They also aid cell division.

The actin cortex form a 2D network providing support for the membrane & allowing transport of protein cargo.

The intermediate filaments are passive rope-like filaments ~~at~~ made from keratin. They anchor to cell junctions & link adjacent cells.

(b)  
(i) The  $\text{Na}^+/\text{K}^+$  has 3 high affinity  $\text{Na}^+$  binding sites in the E1 configuration & 2 low affinity  $\text{K}^+$  sites

4 b  
(i)

In its E1 conformation, the  $\text{Na}^+/\text{K}^+$  ATPase has three high-affinity Na-binding sites and two low-affinity K-binding sites accessible to the cytosolic surface of the protein. The  $K_m$  for binding of  $\text{Na}^+$  to these cytosolic sites is 0.6 mM, a value considerably lower than the intracellular Na concentration of  $\approx 12$  mM; as a result,  $\text{Na}^+$  ions normally will fully occupy these sites. Conversely, the affinity of the cytosolic K-binding sites is low enough that  $\text{K}^+$  ions, transported inward through the protein, dissociate from E1 into the cytosol despite the high intracellular K concentration. During the E1  $\rightarrow$  E2 transition, the three bound  $\text{Na}^+$  ions become accessible to the exoplasmic face, and simultaneously the affinity of the three Na-binding sites becomes reduced. The three  $\text{Na}^+$  ions, transported outward through the protein and now bound to the low-affinity  $\text{Na}^+$  sites exposed to the exoplasmic face, dissociate one at a time into the extracellular medium despite the high extracellular Na concentration. Transition to the E2 conformation also generates two high-affinity  $\text{K}^+$  sites accessible to the exoplasmic face. Because the  $K_m$  for  $\text{K}^+$  binding to these sites (0.2 mM) is lower than the extracellular  $\text{K}^+$  concentration (4 mM), these sites will fill with  $\text{K}^+$  ions. Similarly, during the E2  $\rightarrow$  E1 transition, the two bound  $\text{K}^+$  ions are transported inward and then released into the cytosol.

(ii)

Oouabain binds to the exoplasmic domain of the  $\text{Na}^+/\text{K}^+$  ATPase & inhibits its activity. Thus, animal cells treated with Oouabain cannot maintain the  $\text{K}^+$  &  $\text{Na}^+$  ion gradients.

(ii)

Overall per ATP molecule hydrolyzed the pump moves 3  $\text{Na}^+$  ions & 2  $\text{K}^+$  ions. Increasing the ATP concentration will allow more pump activity.